

USE OF CHEMILUMINESCENT LABELED PROBES FOR FORENSIC AND PATERNITY DETERMINATIONS.

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Introduction.

Since the middle 1980's, issues involving human identification in forensic or paternity cases have been addressed by RFLP DNA analysis using probes radiolabeled with P^{32} (Kanter et al., 1986; Guisti et al., 1986; Baird et al., 1986). The main disadvantage of these types of probes are their short shelf life and the general inconvenience associated with working with radioactive materials. For the past several years our laboratory has been involved in the conversion of these probes to a non-isotopic format. This has been accomplished through the development of oligonucleotide probes labeled with alkaline phosphatase (AP-probe) that recognize VNTR loci (Baum et al. 1990, Neuweiler et al. 1992). The final detection of AP-probes is based on the detection of chemiluminescence with the substrate, adamantyl 1,2-dioxetane phenyl phosphate (LumiphosTM). This detection system offers equivalent sensitivity to P^{32} allowing detection of genomic DNA in the ng range.

Materials and Methods

Standard RFLP methodology was utilized to prepare, digest, size separate and transfer DNA. Starting with DNA bound to dry nylon membranes, the general procedure was as follows:

- Wet Membranes (5 to 10 min at room temp.)
- Hybridization (20 min at 55 °C)
- Wash I (2 times for 10 min each at 55 °C)
- Wash II (2 times for 10 min at 55 °C)
- Rinse with Quick-LightTM Buffer (4 x 1 min dip)
- Spray substrate (Lumi-PhosTM)
- Expose to X-ray film (O.N. at room temp. or 1-6 hrs at 37 °C)

Results and Discussion

Numerous VNTR loci were tested for their utility for DNA typing and for the properties of their respective AP-probes. Each AP-probe was synthesized as an oligonucleotide and directly conjugated with AP. Table 1 provides a list and brief description of the properties of 14 loci available in our laboratory for paternity and/or forensic DNA profiling. Where appropriate, databases have been developed with Hae III and/or Pst I digested DNA from unrelated North American Blacks and Caucasoids individuals. In addition, for some loci, databases have been generated from 14 different ethnic groups (Balazs, 1993).

Table 1. Properties of AP-probes used for the analysis of VNTR loci.

| Locus | SIZE RANGE OF DNA FRAGMENTS (KB) | | | % Heterozygosity |
|--------|----------------------------------|------------------------------|-----------|------------------|
| | Hae III | Restriction Enzyme Hinf I | Pst I | |
| D1S7 | 1.6 - 23 | 0.8 - 25 | | 99 |
| D1S339 | 0.6 - 12 | 1.2 - 13 | 1.2 - 13 | 95 |
| D2S44 | 0.2 - 11 | 1.5 - 10 | 6.8 - 16 | 96 |
| D2S92 | 2.4 - 13 | | | 94 |
| D4S163 | 1.3 - 16 | 1.2 - 12 | 3.6 - 16 | 95 |
| D6S132 | 1.0 - 8.5 | 4.3 - 12 | 1.7 - 9 | 92 |
| D7S21 | | 3.0 - 20 | 4.5 - 20 | 97 |
| D7S467 | 1.6 - 7 | | 3.5 - 15 | 94 |
| D10S28 | 0.2 - 13 | | | 94 |
| D12S11 | | 3.0 - 18 | 4.7 - 25 | 95 |
| D16S85 | 0.4 - 7 | 0.3 - 9 | 0.6 - 10 | 92 |
| D17S79 | 0.6 - 2.4 | 1.0 - 4.3 | 2.5 - 5.7 | 88 |
| D18S27 | | 1.0 - 6.2 | 4.0 - 9 | 92 |
| DNF24 | | 1.0 - 23 | 2.0 - 25 | multilocus |

The use of AP labeled probes has a number of advantages over traditional radiolabeled probes. These advantages include:

1. Stability

AP-labeled probes are stable for at least one year.

2. Safety

AP-labeled probes do not require containment or monitoring because they are not radioactive. The reagents utilized do not require special handling.

3. Hybridization

The hybridization conditions are generally milder (55 °C vs 65 °C) and shorter (20 min vs several hours) resulting in a decreased loss of DNA from filter. Exposures to the X-ray film are at room temperature or 37 °C instead of

70 °C. Because light emission lasts for up to 3 days, re-exposures are easily obtained. The resulting images are sharper than those obtained with ³²P-labeled probes, therefore AP-probes give better resolution and greater power of exclusion.

4. Speed

Because of the fast and simple protocol it is possible to obtain at least one set of results per day. By doing daily rehybridizations with various AP-probes, it is easy to produce results with four different probes in a one week period.

5. Rehybridization

The AP-labeled probe is inactivated by heating at 65 °C for 20 min.

Hybridization of membranes can be readily interchanged between AP-probes and radioactive probes. Therefore, rehybridization can be performed with ³²P-labeled or AP-labeled probes.

In summary, the use of AP-labeled probes is faster, safer, and easier than radiolabeled probes. They are ideal for laboratories that want to move away from the use of radioactive materials and/or without the resources for radiolabeled containment or for laboratories interested in fast turnaround time.

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