

## HUMAN RED CELL ACID PHOSPHATASE (ACP1): GENETIC TYPING AT THE DNA LEVEL

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Human red cell acid phosphatase (ACP1) is one of the best known of the forensically useful genetic polymorphisms. Two common alleles - ACP1\*A and ACP1\*B - are found in all major population groups. A third allele - ACP1\*C - is common in Caucasians and a fourth - ACP1\*R - is common in Blacks of African origin. A number of rare alleles have also been noted.

Each of the common alleles is expressed as a pair of electrophoretically distinct isozymes, termed fast (**f**) and slow (**s**) according to their anodal mobility. The **f**:**s** isozyme ratios for the \*A, \*B, and \*C alleles are 2:1, 4:1, and 1:4 respectively [1]. This allelic variation in **f**:**s** ratio accounts for observed phenotypic variation in activity modulation by purines and folates, and in phosphotransferase activity [2-9]. The allele dependent variation in the amount of enzyme produced in red cells is independently regulated.

Recent protein and DNA sequencing studies show that the **f** and **s** isozymes are identical over the protein sequence regions spanning residues 1-39 and 74-157 but differ significantly (59%) in sequence in the middle region, residues 40-73 [10-12]. The sequence differences in this middle "signature" region presumably account for the differences in the catalytic and molecular properties of the **f** and **s** isozymes. The **f** and **s** isozyme pairs have been postulated to be generated by an alternative splicing mechanism in which the **f** and **s** signature sequences are encoded in mutually exclusive exons [10,13]. The gene structure of the ACP1 locus is consistent with this hypothesis [14].

Gene sequence analysis of the \*A, \*B, and \*C alleles was undertaken to identify base substitutions that distinguish between them. A base substitution at codon position 105 should account for the glu-arg amino acid substitution that distinguishes \*A from \*B and \*C [11,15]. Other base differences should account for the allele dependent variation in **f**:**s** isozyme ratio and in red cell activity level. Base differences between the alleles can also be used for general typing purposes.

### ACP1 SEQUENCE AND GENE STRUCTURE

Genomic sequence spanning approximately 6 Kb of the ACP1 locus has been determined. Six linearly positioned exons have been identified; these account for all the coding sequence except for amino acid residues 1-13 at the N-terminus. The missing coding sequence appears to be located in an exon or exons at least 4 Kb upstream of the first sequenced exon. Genomic sequences of homozygous \*A, \*B, and \*C types show the same gene structure.

As previously predicted, the **f** and **s** signature sequences are encoded in different exons, exons 3F and 3S; the two exons are of equal length. The intervening sequence is 41 bp, too short to serve as a functional intron. This gene structure is consistent with the mutually exclusive alternative splicing hypothesis.

The \*A, \*B, and \*C sequences were found to differ at three sites as indicated in the table below:

	<u>*A</u>	<u>*B</u>	<u>*C</u>
Exon 3F base 15	C	C	T
Exon 3S base 12	T	C	C
Exon 5 base 24	G	A	A

The first two substitutions are silent but may play a role in regulating f:s ratios. The substitution in exon 5 is in the second base of codon 105 and accounts for the Glu-> Arg difference that distinguishes the \*A type from the \*B and \*C types.

#### DETECTION OF \*A, \*B, AND \*C ALLELES

The base substitution in exon 5 creates a TaqI restriction site in the \*A allelic sequence not present in the \*B and \*C allelic sequences; this has been exploited in a typing system to distinguish the \*A allele from the \*B and \*C alleles [15]. Amplification using primers ACP38 (5'GGATGTTTCAGAAGACCCTAGCAG3', sense strand) and ACP18 (5'GCTCCCAAGTAGTTCAATTTAGC'3) yields a 149 bp product for each of the \*A, \*B, and \*C alleles. Restriction with TaqI cleaves the \*A allele product into 106 bp and 43 bp fragments; the \*B and \*C allele products are not cut.

The base substitution in exon 3F creates a CfoI restriction site in the \*A and \*B allelic sequences not present in the \*C allelic sequence; this can be exploited in a typing system to distinguish the \*C allele from the \*A and \*B alleles. Amplification using primers ACP10 (5'GTCGATCACCCATTGCAGAAGC3', sense strand) and ACP44 (5'CACACAACCTCAAGTCCAAGGACG3') yields a 303 bp product for each of the \*A, \*B, and \*C alleles. Restriction with CfoI cleaves the \*A and \*B allele products into 177 bp and 126 bp fragments; the \*C allele product is not cut.

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